



Bio dechlorization of Some Industrial dyes by use *Pencillium expansum* and *Aspergillus niger*

Hada Abas Mohammed

Environmental Research Center- Babylon University

Corresponding author: jasimsalman@uobabylon.edu.iq

To cite this article:

Mohammed. H. A, Bio dechlorization of Some Industrial dyes by use *Pencillium expansum* and *Aspergillus niger*. *Mesop. environ. j.*, 2016, Vol.3, No.1;37-44.

This work is licensed under a [Creative Commons Attribution-Non Commercial-No Derivatives 4.0 International License](https://creativecommons.org/licenses/by-nc-nd/4.0/).



Abstract

This research is interested in using the two species of fungi such as *Penicillium expansum* and *Aspergillus niger* for Biodegradation of Safranin dye as industrial and Carcinogenic dyes, The results showed that *P. expansum* was the more efficient from *Asp. niger* in Biodegradation of the Safranin dye. The percentage of Bio decolonization about 41.035% during incubation at 120 hours, while when use *Asp. niger* the percentage of Bio decolonization was about 38.952% during same time of incubation.

Keywords; fungi, Safranin; biodecolorization, pollution.

Introduction

Safranines are the azonium compounds of symmetrical 2,8-dimethyl-3,7-diamino-phenazine. They are obtained by the joint oxidation of one molecule of a para-diamine with two molecules of a primary amine; by the condensation of para-aminoazo compounds with primary amines, and by the action of para-nitrosodialkylanilines with secondary bases such as diphenylmetaphenylenediamine [1].

Biosurfactants are structurally and functionally diverse amphiphilic, surface active compounds which lower the surface and interfacial tension between individual molecules at respective surfaces and interfaces. Thus, these are very important in the living systems and can be regarded as the backbone of the biological membranes which promise the transport and exchange of the various important materials [2,3]. Biosurfactants are ecologically safe and can be applied in bioremediation processes. The microorganisms which produce biosurfactants can also be used in the various bioremediation technologies like solubilisation and removal of oil from contaminated soil, sludge in oil storage tank etc [4].

Synthetic dyes are coloring agents mainly used in textile industries which generate a huge amount of waste water in the process of dying. It is estimate that these industries discharge around 28,000 tons of dyes worldwide every year in the environment [5].and variety of bacteria and few fungi are reported to produce biosurfactants using renewable sources [6,7].

In recent years, many fungi used in many of the studies and intensively on removing dyes from wastewater [6]. From the textile industry can get a large amount of liquid waste that can cause serious environmental problems. It is estimated that the liberation of 10-15% of dyes in water treatment [7] is responsible for the color dyes that affect the photosynthetic activity of aquatic life by reducing the intensity of light propagation may also be toxic to some aquatic animals and plants due to the presence of aromatic substances, and metals, chlorides [8].

There are many factors that affect the ability and speed of fungus toxin analysis. The first is the physical properties or formalism of hydrocarbons. In general, the contaminants statistics molecular structure is broken and easier analysis of contaminants with complex structure. Second factor, which affects the speed of cracking pollutants, is the temperature. The degree low temperature leads to slow the process of analyzing contaminants while the high temperature to some extent speed up the process. Fungi prefer acidic environment on a neutral environment. And oxygen is an important element in the fungal metabolism processes. As the first step in the analysis of hydrocarbons is to add oxygen to it, and thus the lack of oxygen in the environment leads to the slow process of analysis [9].

It is clear that micro-communities in the environment works hand in hand with fungi to analyze and break down pollutants, and converted into carbon dioxide and water. The fungus that decomposition of the wood of the most effective types of aromatic pollutants in the analysis of (toxic components of oil), as well as some of the materials and compounds ,In addition to some of the materials and compounds added to it elemental chlorine (such as some types of pesticides that are difficult to get rid of them) [10].

If it is not controlled in the biological treatment process, it is possible not to break down organic pollutants occur fully, leading to obtain products more toxic and that can have a greater ability to move from primary pollutants[11].

In This research examination of the fungi toward biodegradation of as industrial and Carcinogenic dyes such as Safranindye by used the two species of fungi were *P.expansum* and *Asp.niger*.

Materials and Methods

Preparation the fungal inoculums

When isolate *P.expansm* from *P.expansm* isolate from fruits and vegetables, and *Asp.niger* was isolated from air, and after this incubated fungal isolates incubator for seven days at a temperature of 25° C and after the incubation period of fungal isolates were purified on the PDA media and incubated for seven days at a temperature of 25 ° C.

After incubation, the fungal inoculum was attended the audacity to take disk of a fungal culture seven days old by using a cork borer diameter of 5 mm and add 10 ml of sterile distilled water to the fungal disk [12].

Preparation Dye Solution

The Safranin (Chemical formula= $C_{20}H_{19}ClN_4$, Formula weight= $350.85 \text{ g}\cdot\text{mol}^{-1}$) supplied by BHD Chemicals (Figure 1,2). The solution of Safranin were prepared by dissolving appropriate amounts (accurate weighed) of dry powdered dye in double distilled water to prepare Stock solution (1000 mg L^{-1}). The experimental solution was obtained by dilutions were made to obtain the working solution at desired concentrations [13].

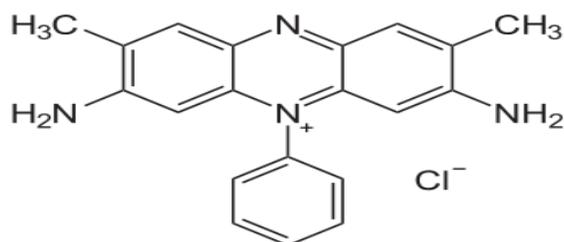


Fig. 1: The chemical structure of Safranin dye



Fig. 2: Safranin in aqueous solution

Experimental

Effect of speciesfungion fungi species on the Dye

Add The fungal inoculum Previously the record to 100ml of Safranin solution was measured and added to the content in each conical flask. The content was shaken rigorously and continuously for 30,60,90,120,150, 180, 210, 240, 270, and 300 min(delete) respectively what?. The particles of the adsorbent was separation by centrifuged from solution to obtain the equilibrium concentration.

The final concentration of Safranin read was estimated for each sample spectrophotometrically at the wavelength corresponding to maximum absorbance for Safranin ($\lambda_{\text{max}}=530\text{nm}$) using a spectrophotometer (UV/VIS-Jenway ,6800, German) as in the Figure 1. A graph of removal Safranin read percentage (g/L) versus time (hour) was plotted for Safranin. Generally the amount of dye removal was calculated from following equation:

$$\text{Biodecolorization\%} = (A^{\circ} - A) / A^{\circ} \times 100$$

A° and A is the absorbance of concentration of dye before and after Biodecolorization respectively[13, 14].

Results and Discussion

The results of this study show that more efficient use of *P.expansum* in Biodecolorization of Safranin dye from the use of *Asp. niger*. The results showed that the amount of absorbance decreases with time treatment, so that the concentration of dye is decrease. When used *P.expansum* the absorption of Safranin during 120 hours of treatment become 0.2892 compared with control while the absorption when used *Asp. niger* at 120 hours about 0.5543 (figure 3). And then change in the color of the dye from red to colorless gradually has been observed when using *P.expansum* and *Asp.niger*. It was rated assessment of the deterioration and decolourization as the disappearance of the color of the plate Petri, through the development of fungal mycelium. Safranin dye has utilized to give the positive result and biodegradable fungus was showed. The color of this dye has turned to pale, finally the current yellow zone around the mycelium. The researchers results showed a small part of the some dyes have also accumulated used mushrooms fungi(all mushrooms word converted into fungi or fungus in all research), mycelium have turned to red[15].

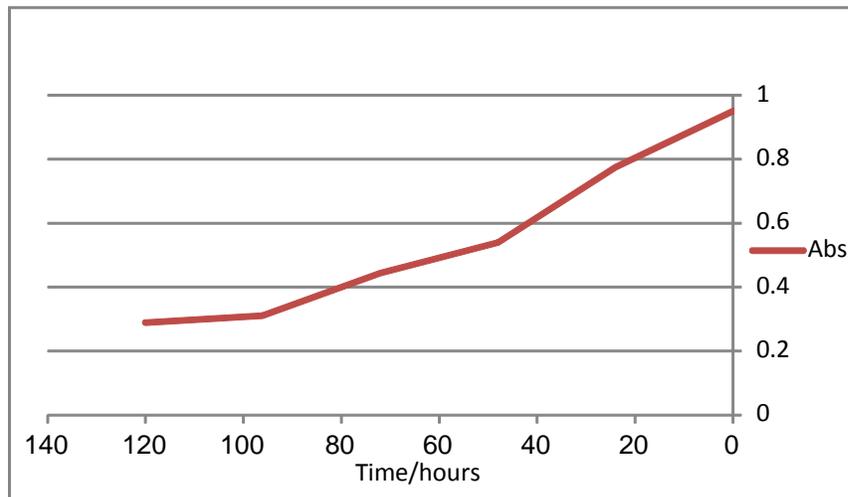


Fig.3: The absorption of Safranin by used *P.expansum*.

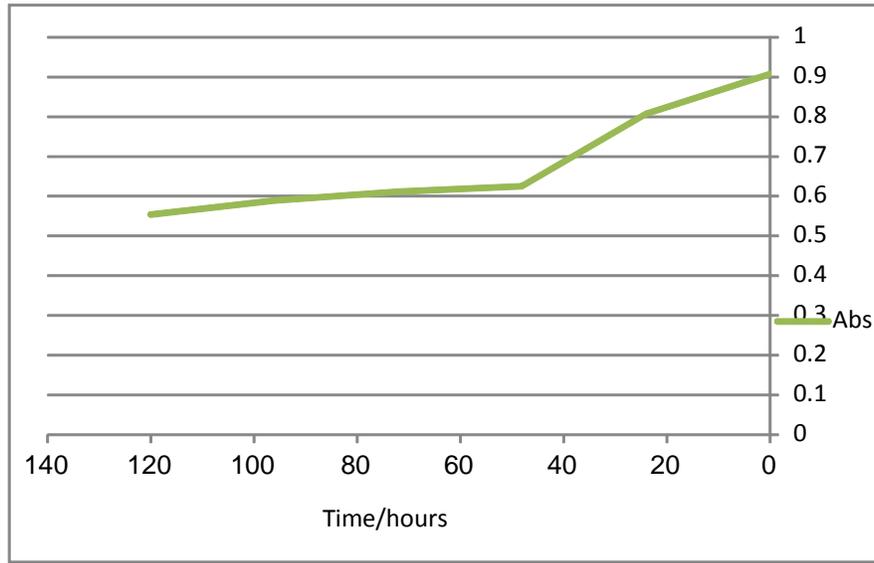


Fig.4: The absorption of Safranin by used *Asp. niger*

After using *P.expansum* was removal percentage of dye Safranindye about 37.135% at 120 hours from treatment compared with removal of dye on the first day of treatment using the same fungus it's about 20.708% (Figure 5, Table 1), In recent years, fungi and bacteria used as an application or technique to remove or absorbing dyes that form risk on a health [16,17,18] ,While Biodegradation percentage of Congo red dye by using *Asp.niger* at 120 hours about 38.952% compared with exposing the dye to the same fungus during the first 24 hours of treatment it's about 17.035% (figure 6, Table 1). Others Fungal such as *Phlebiatremellus*, *Phanerochaetechrysosporium*, *Tranmetesversicolor*, *Fusariumoxysporum*, *Aspergillusflavus* and *Trichodermaviride* are also able to removes the dyes, and most research have been limited to the decolourization of a one dye or even to mixtures of dyes. Nevertheless, a biodecolourization system must sustain its ability upon exposure to real wastewater conditions[19].This study is consistent with another study, explained the role of fungi in the removal of carcinogenic chemical dyes[20].

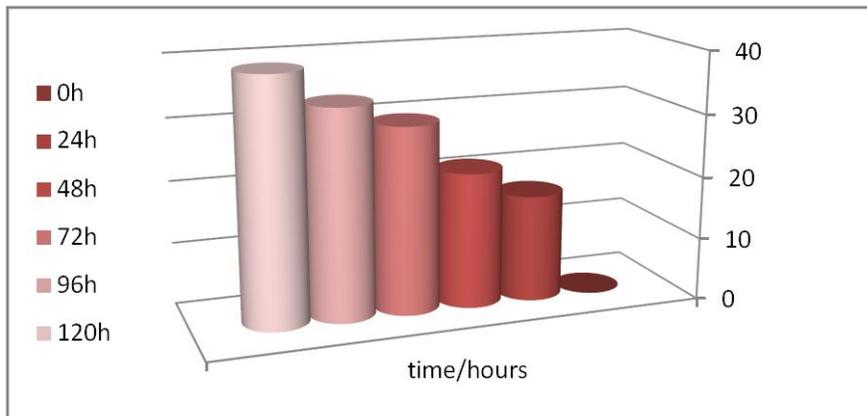


Fig.5: percentage of bio decolonization to Safranin dye treated by *Penicillium expansum*.

Table 1: percentage of bio decolonization to Safranin dye treated by *Penicilliumexpansum*, *Aspergillusniger*.

Time/hours	percentage of Biodecolorization by fungi	
	<i>Penicilliumexpansum</i>	<i>Aspergillusniger</i>
24 h	20.708	17.035
48 h	25.831	21.630
72 h	32.678	29.802
96h	37.135	33.416
120h	41.035	38.952

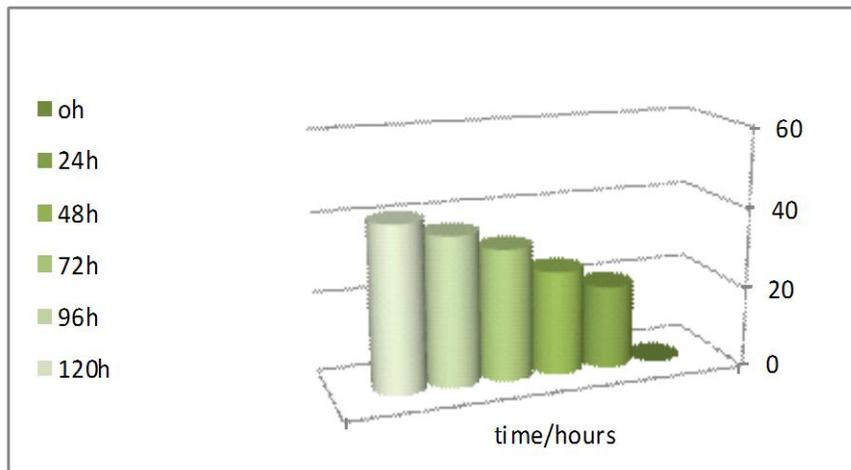


Fig.6: percentage of bio decolonization to Safranin dye treated by *Aspergillusniger*

Reference

- [1] "Safranin O" (PDF). LabChem.Retrieved 10 March 2016.
- [2] Lang S. Biological amphiphiles (microbial biosurfactants). *Curr Opin Colloid Interphase Sci* 7: 12-20.2002.
- [3] Satpute SK, Banpurkar AG, Dhakephalkar PK, Banat IM, Chopade BA. Methods for investigating biosurfactants and bioemulsifiers: a review. *Crit Rev Biotechnol* 30: 127-144.2010.
- [4] Banat IM. Biosurfactants production and possible uses in microbial enhanced oil recovery and oil pollution remediation: A review. *Bioresour Technol* 51: 1-12.1995.
- [5] Bras, A.; Gomes, A.; Ferra, M.I.; Pinheiro, H.M. and Goncalves, I.C. Monoazo and diazo dye decolourization studies in a methanogenic UASB reactor. *J. Biotechnol.*, 115, 57- 66 .2005.
- [6] Satpute SK, Banat IM, Dhakephalkar PK, Banpurkar AG, Chopade BA. Biosurfactants, bioemulsifiers and exopolysaccharides from marine microorganisms. *Biotechnol Adv* 28: 436-450.2010.
- [7] Patil JR, Chopade BA .Studies on bioemulsifier production by *Acinetobacter* strains isolated from healthy human skin. *J Appl Microbiol* 91:290-298.2001.
- [8] Dhaneshvar, N.; Ayazloo, M.; Khatae A.R. and Pourhassan M., 2007. Biological decolourization of dye solution containing malachite green by microalgae *Cosmarium* sp. *Bioresour Technology*. 29.1-7.2003.
- [9] Singh, Harbhajan. *Mycoremediation: fungal bioremediation*. New York: Wiley-Interscience. ISBN 0-471-75501-X. 2006.
- [10] Thomas, S.A. *Mushrooms: Higher Macrofungi to Clean Up the Environment*, Battelle Environmental Issues, Fall 2000
- [11] Sharma, H. D., & Reddy, K. R. "Geoenvironmental Engineering". Hoboken, New Jersey: John Wiley & Sons, Inc. 2004.
- [12] Gopi, V; Ugrade, A; and Soundararajan N. Bioremediation potential of individual and consortium Non-adapted fungal strains on Azo dyes containing textile effluent. *Advances in applied Science Research* 3(1):303-311.2012.
- [13] Alejandro. B; Rodri'guez-Navarro; Christiane Cabral de Melo B. Nelson Batista; Nilton Morimoto; Pedro Alvarez-Lloret; Miguel Ortega-Huertas; Victor M. Fuenzalida; Jose. I; Arias Juan. P; Wiff Jose. L. Arias" Microstructure and crystallographic-texture of giant barnacle (*Austromegabalanus psittacus*) shell" *Journal of Structural Biology* 156, 355-362. 2006.
- [14] Salman ,J.M.; Amrin, A.R.; Hassan, F.M.; Jouda, S.A. Removal of congo red dye from aqueous solution by using natural materials. *Mesop. environ. j.* 1, 3. 82-89, 2015.
- [15] Gupta M. and Manisha K. Biodegradation of textile dye Congo red by Fungus *Mucormuced*. *Indian Journal of Fundamental and Applied Life Sciences*. 2. 2.251-255.2012.

[16] **Ayed L, Mahdhi A, Cheref A, Bakhrouf A:** Decolorization and degradation of azo dye Methyl red by an isolated *Sphingomonas paucimobilis*: Biototoxicity and metabolites characterization. Desalination .274.272–277, 2011.

[17] **Saratale, R. G.; Saratale, G. D.; Chang, J. S.; Govindwar, S. P.** Bacterial decolorization and degradation of azo dyes: A review. J Taiwan Inst Chem Eng .2, 138–157, 2011.

[18] **Dawkar, V.V.; Jadhav, U.U.; Jadhav, S.U. and Govindwar, S.P.** Biodegradation of disperse textile dye brown 3REL by newly isolated *Bacillus* sp., VUS. J Appl Microbiol .105,.14–24, 2008.

[19] **Kannahiand, M. and Uma, N.** Decolourization of textile dye using *Aspergillus flavus* and *Trichoderma viride* . Res. Environ. Life Sci, 4,.115-118, 2011.

[20] **Mohammed, H. A.** Biodecolorization of Congo red Dye by using two species of Fungi. Mesop. environ. j., 1, 4, 50-56. 2015.